



RESEARCH ARTICLE

Exploratory analysis of the proteomic profile in plasma in adults with Down syndrome in the context of Alzheimer's disease

Olivia Wagemann^{1,2}  | Georg Nübling¹ | Francisco Jesús Martínez-Murcia³ | Elisabeth Wlasich¹ | Sandra V. Loosli^{1,4} | Katja Sandkühler¹ | Anna Stockbauer^{1,2} | Catharina Prix¹ | Sabrina Katzdobler^{1,2} | Agnese Petrera⁵ | Stefanie M. Hauck⁵ | Juan Fortea^{6,7,8} | Rocío Romero-Zalíz^{3,9} | Carmen Jiménez-Mesa³ | Juan M. Górriz Sáez³ | Günter Höglinger^{1,2,10} | Johannes Levin^{1,2,10} 

¹Department of Neurology, University Hospital, Ludwig-Maximilians-University (LMU) Munich, Munich, Germany

²German Center for Neurodegenerative Disease (DZNE), Munich, Germany

³Department of Signal Theory, Telematics and Communications, Andalusian Institute in Data Science and Computational Intelligence (DaSCI) at University of Granada, Granada, Spain

⁴Department of Neurology, University Hospital Zurich, Zurich, Switzerland

⁵Metabolomics and Proteomics Core, Helmholtz Zentrum München, German Research Center for Environmental Health (GmbH), Neuherberg, Germany

⁶Sant Pau Memory Unit, Hospital de la Santa Creu i Sant Pau - Biomedical Research Institute Sant Pau, Barcelona, Spain

⁷Centro de Investigación Biomédica en Red de Enfermedades Neurodegenerativas, CIBERNED, Madrid, Spain

⁸Barcelona Down Medical Center, Fundació Catalana Síndrome de Down, Barcelona, Spain

⁹Information and Communication Technologies Research Centre (CITIC-UGR), University of Calle Periodista Rafael Gómez Montero, Granada, Spain

¹⁰Munich Cluster for Systems Neurology (SyNergy), Munich, Germany

Correspondence

Johannes Levin, Department of Neurology, LMU University Clinic Munich, Marchioninistraße 15, 81377 Munich, Germany.
Email: johannes.levin@med.uni-muenchen.de

Abstract

INTRODUCTION: Adults with Down syndrome (DS) show increased risk for Alzheimer's disease (AD) due to the triplication of chromosome 21 encoding the *amyloid precursor protein* gene. Further, this triplication possibly contributes to dysregulation of the immune system, furthering AD pathophysiology.

METHODS: Using Olink Explore 3072, we measured ~3000 proteins in plasma from 73 adults with DS and 15 euploid, healthy controls (HC). Analyses for differentially expressed proteins (DEP) were carried out, and pathway and protein network enrichment using Gene Ontology, Kyoto Encyclopedia of Genes and Genomes (KEGG), and STRING database was investigated. Within DS, the LASSO (least absolute shrinkage and selection operator) feature selection was applied.

Funding information: Alzheimer Forschung Initiative, Grant/Award Number: #22049; Deutsche Forschungsgemeinschaft; Fuse Deutsche Forschungsgemeinschaft; Munich Cluster for Systems Neurology, Grant/Award Numbers: EXC 2145, 390857198; Bundesministerium für Bildung und Forschung project CLINSPECT-M, Grant/Award Numbers: FKZ161L0214B, FKZ161L0214C, FKZ161LW0248; Else Kröner-Fresenius-Stiftung, Grant/Award Number: 2020_EKEA.09; Jerome LeJeune Foundation; Verum Foundation; Fondo de Investigaciones Sanitarias, Carlos III Health Institute, Grant/Award Numbers: INT21/00073, PI20/01473, PI23/01786; Centro de Investigación Biomédica en Red sobre Enfermedades Neurodegenerativas Program 1; Unión Europea; Una Manera de Hacer Europa; National Institutes of Health, Grant/Award Numbers: R01 AG056850, R21 AG056974, R01 AG061566, R01 AG081394, R61AG066543; Department de Salut de la Generalitat de Catalunya, Pla Estratègic de Recerca i Innovació en Salut, Grant/Award Number: SLT006/17/00119; Fundación Tatiana Pérez de Guzmán el Bueno, Grant/Award Number: IIBSP-DOW-2020-151; Horizon 2020–Research and Innovation Framework Programme from the European Union, Grant/Award Number: H2020-SC1-BHC-2018-2020

This is an open access article under the terms of the [Creative Commons Attribution](https://creativecommons.org/licenses/by/4.0/) License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

© 2025 The Author(s). *Alzheimer's & Dementia* published by Wiley Periodicals LLC on behalf of Alzheimer's Association.

RESULTS: We identified 253 DEP between DS and HC and 142 DEP between symptomatic and asymptomatic DS. Several pathways regarding inflammatory and neurodevelopmental processes were dysregulated in both analyses. LASSO feature selection within DS returned 15 proteins as potential blood markers.

DISCUSSION: This exploratory proteomic analysis found potential new blood biomarkers for diagnosing DS-AD in need of further investigation.

KEYWORDS

Alzheimer's disease, biomarker, Down syndrome, neuroinflammation, plasma

Highlights

- Inflammatory pathways are dysregulated in symptomatic versus asymptomatic DS.
- NFL and GFAP are confirmed as powerful biomarkers in DS with clinical and/or cognitive decline.
- Further circulating proteins were identified as potential blood biomarkers for symptomatic DS.

1 | BACKGROUND

In Alzheimer's disease (AD), amyloid-plaques and neurofibrillary tau tangles (NFT) make up the pathophysiological hallmarks causing neurodegeneration,^{1,2} known to cause decline in cognitive capacity and everyday functioning. However, other processes like vascular dysfunction, oxidative stress, endosomal and lysosomal dysfunction, and neuroinflammation are likely to play a pivotal role as well.^{3,4}

Adults with Down syndrome (DS) present with a triplication of the *amyloid precursor protein (APP) gene*, due to the presence of three copies of chromosome 21 (chr21),⁵ which results in the early accumulation of intracellular amyloid already in childhood,⁶ eventually leading to extracellular accumulation of amyloid, the formation of solid amyloid-plaques, and subsequent intracellular tau phosphorylation, ultimately resulting in progressive cognitive impairment.⁷

Even though DS-AD largely parallels the pathophysiological and clinical presentation of AD in the carriers of the autosomal dominant gene mutations in *presenilin 1 (PSEN1)*, *presenilin 2 (PSEN2)*, or *APP*, the diagnosis of dementia due to DS-AD remains challenging, predominantly due to the heterogeneous presentation of clinical symptoms, widely varying level of baseline cognitive capacity and the lack of standardized tests to assess and evaluate these.^{8,9}

Therefore, diagnosis often relies on external anamnesis since those affected may not recognize or prioritize memory decline as a significant issue. Behavioral changes or loss of daily living skills are more frequently reported, but these symptoms can be highly variable.^{10,11} The lack of awareness among families, caregivers, and clinicians exacerbates these diagnostic challenges,¹² which can lead to a significant delay of the diagnosis.

A lot of effort has been put into researching biomarkers in DS-AD by assessing AD pathology in brain imaging, cerebral spinal fluid (CSF), and blood. Considering cost-effectiveness, accessibility in a clinical

setting, and the minimally invasive nature of sample obtainment, blood-based biomarkers are of high interest, potentially mirroring AD-related changes of the brain in DS within in the peripheral bloodstream.¹³

Levels of A β 42 in plasma have been reported to be elevated in DS, yet reports on the diagnostic and prognostic performance of both A β 42 and A β 42/40 ratio remain inconsistent.^{14–18} Markers of tau pathology, however, namely pTau-181 and pTau-217, reliably differentiate between symptomatic and asymptomatic DS and have been reported to correlate well with tau and amyloid pathology as assessed by positron emission tomography (PET).^{14,19–21} Further, plasma levels of neurofilament light chain (NFL), a marker of axonal degeneration, rise early and correlate well with cognitive decline²² as well as amyloid load and neurodegeneration in brain imaging, offering strong diagnostic utility.^{15,23} Finally, glial fibrillary acidic protein (GFAP), relating to astrocytic activity, rises as early as in the third decade, correlates well with cognitive status as well as tau and amyloid burden in PET, and shows good performance for predicting disease progression, which interestingly could not be replicated in CSF.^{21,24} Recently, we have also shown that the presynaptic marker beta-synuclein provides good discriminatory power in DS-AD, rising even before pTau-181.²⁵

Despite the genetic basis of DS-AD, resulting in a predictable sequence of pathophysiological events,⁵ significant variability in symptom onset and presentation remains, warranting the investigation of additional pathophysiological processes that might be directly or indirectly caused by increased levels of proteins originating from chr21.^{26–29} A neuroinflammatory phenotype in DS-AD, that is, distinct from sporadic AD (sAD) has been suggested²⁷ and neuropathological studies reported a neuroinflammatory protein profile evolving across the DS-AD continuum.²⁶ In blood, differences in circulating inflammatory proteins compared to euploids have been identified, in line with chronic autoinflammation possibly contributing to DS-AD²⁹ while another study, targeting 20 inflammation-related proteins,

further found differences in DS-AD and mildly impaired DS compared to asymptomatic DS with excellent accuracy,³⁰ underscoring the need to further investigate interconnected processes through exploratory proteomic approaches.

We aimed to explore the proteomic profile of individuals with DS using OLINK technology to identify proteins and pathways influencing AD progression thereby leveraging a broader, exploratory approach. The findings may uncover biomarkers and therapeutic targets, providing a foundation for future hypothesis-driven research into DS-AD pathophysiology.

2 | METHODS

2.1 | Participants and clinical workup

All participants included in this investigation are either part of our AD21 cohort study or our study for biological samples in neurodegeneration (Biobank für translationale Neurodegeneration), and were recruited from the outpatient clinic at LMU University Hospital Munich in Germany. The former, AD21, investigates AD in adults with DS, using clinical characterization, neuropsychology, neuroimaging, and biofluid analysis with annual follow-up visits. Further information on the study design as well as inclusion and exclusion criteria can be found in the supplements (Table S1). The latter study provided the euploid healthy controls (HC) included in this analysis and aims at collecting biofluids and basic demographic information within the framework of a biobank. Included in this analysis were HC from this study where clinically as well as by patient account there was no sign of cognitive decline and no hint at cognitive symptoms impacting daily living activity and further no diagnosis of cognitive impairment of any kind, known structural brain lesions, or the diagnosis of a neurodegenerative disease.

Each individual or their respective legal proxy provided informed written consent prior to inclusion. Both studies are approved by the LMU ethics committee (DS: #17-126, HC: #20-0997) and conducted in accordance with the Declaration of Helsinki.

Our study sample reflects the demographics of adults with DS in Germany, where participants were predominantly of European descent, specifically White/Caucasian. While ethnic diversity within this cohort is limited, we sought to ensure inclusivity by recruiting participants from various age groups as well as intellectual disability.

Chromosome analysis in DS was conducted to assess the accurate type of trisomy 21 (full, translocation, mosaicism) where possible. Intellectual disability (ID) was stratified according to Diagnostic and Statistical Manual of Mental Disorders, 5th edition (DSM-V) criteria³¹ into mild, moderate, severe, or profound, based on the individuals' best-ever level of functioning, obtained from interviews with caregivers, neuropsychological assessment, behavioral observations, and review of medical records.

Symptomatic diagnosis was reached independently by two neurologists referencing a predefined diagnostic algorithm⁹: changes in cognition, behavior, and activities of daily living were assessed based on patient and caregiver information. Subsequently, differential diag-

RESEARCH IN CONTEXT

- 1. Systematic review:** The authors conducted a comprehensive review of the existing literature leveraging established databases (e.g., PubMed, Google Scholar). There were very few publications using proteomics to explore blood samples in Down syndrome (DS), especially in the context of Alzheimer's disease (AD) or neuroinflammation, those uncovered are cited accordingly in the manuscript. We identified what we consider critical gaps in proteomic analyses in Down syndrome, setting the stage for this study's proteomic approach using the Olink Explore 3072 platform.
- 2. Interpretation:** Our findings contribute new insights into DS-AD by identifying key pathways associated with immune and neurodegenerative processes as well as providing potential blood biomarkers for AD in the DS population which may serve as early indicators or therapeutic targets.
- 3. Future directions:** Longitudinal analyses are needed to verify the current results and to further investigate the mechanistic pathways linking immune dysregulation with neurodegeneration in Down syndrome.

noses were excluded via neurological, laboratory blood test, and individual cognitive performance assessments performed by trained neuropsychologists using the validated German version of the Cambridge Cognitive Examination for Older Adults with Down Syndrome (CAMCOG-DS).^{8,32} The CAMCOG-DS consists of 45 items and 7 subscales (orientation, language, memory, attention, praxis, abstract thinking, and visual perception). A more detailed description of the specific tasks can be found elsewhere.⁸ This eventually led to the diagnosis of either asymptomatic DS (aDS) or symptomatic DS (sDS), the latter including dementia as well as mild cognitive impairment in the context of DS-AD.

Alzheimer's pathology was further verified referencing the A/T/N criteria,¹ where possible, using validated AD biomarkers such as cerebral spinal fluid (CSF) for assessment of phospho-tau-181 (pTau181, cutoff 61 pg/mL) and A β 1-42/1-40 ratio (A β -ratio, cutoff 5.5%) with the respective Innotest assays. Further, if possible, cerebral imaging was acquired as part of the clinical workup. For this, magnetic resonance imaging (MRI) including T1- (repetition time [TR] 2560 ms, TI: 1100 ms, flip angle: 7°, CAIPIRINHA = 2, field of view [FOV] = 256 mm \times 256 mm, slice thickness 0.8 mm, 224 slices per slab) and T2-weighted sequences (TR 2800 ms, echo time [TE] 405 ms, GRAPPA = 2, FOV = 256 mm \times 256 mm, slice thickness 0.8 mm, 224 slices per slab) as well as a fluid-attenuated inversion recovery sequence (TR 5000 ms, TE 393 ms, GRAPPA = 2, FOV = 256 mm \times 256 mm, slice thickness 1 mm, 176 slices per slab) were obtained using a 3 Tesla Siemens Scanner (Siemens, Erlangen, Germany). Positron emission tomography (PET)

imaging was performed with a Siemens Biograph-64 system. 18F-florbetaben PET (295 MBq) images were acquired 90–110 min after injection. 18F-PI-2620-PET (185 MBq) was performed using dynamic emission recording (0–60 min after injection, 30–60 min window for visual interpretation). For intensity scaling, a cerebellar grey matter reference was used for all tracers.

2.2 | Collection and measurement of plasma samples

Blood samples for both study cohorts were collected into ethylenediaminetetraacetic acid (EDTA) tubes from non-fasting participants, centrifuged at $2000 \times g$ for 10 min to obtain plasma, aliquoted into polypropylene tubes, and subsequently stored at -80°C within 30–45 min after initial blood draw.

For protein analysis, we leveraged the OLINK Explore 3072 Panel (Uppsala, Sweden; Lot #B31341; consisting of the following individual Explore 384 kits: Inflammation Lot #B24609, Inflammation II Lot #B23708, Oncology Lot #B23705, Oncology II Lot #B24546, Cardiometabolic Lot #B23706, Cardiometabolic II Lot #B23710, Neurology Lot #B23707, Neurology II Lot #B23711), a multiplex immunoassay platform designed for human protein biomarker discovery, measuring the concentration of approximately 3000 proteins in one bio sample.³³ For this, blood samples had to be thawed, pipetted on the assay plate, and refrozen before being sent to OLINK for measurement. This proximity extension assay uses highly specific antibodies that bind to their respective protein and hybridize their attached nucleotide single strands with neighboring antibodies on the protein's surface. The resulting double strand is then amplified and quantified via next-generation-sequencing read-out. After quality control, the final data set is obtained as an arbitrary unit of normalized protein expression values (NPX, log₂-transformed). For this run, the intra-assay coefficient of variation (%CV) for the proteins measured in each kit ranged from 6%CV to 13%CV, with a median of 9%CV. The limit of detection (LOD), calculated for each kit individually, ranged from -11.21 to 2.11 .

2.3 | Statistical analysis

All statistical analyses were run on R (version 2023.06.1+524). Exploratory proteomics analysis was conducted using the R packages Olink Analyze, clusterProfiler, glmnet as well as the STRING database website.³⁴

Baseline demographics were summarized as mean \pm standard deviation for continuous variables and count percentage for categorical variables. Group comparisons were conducted via Mann–Whitney *U*-tests for continuous and Fisher's exact tests for categorical variables.

t-Tests investigating differentially expressed proteins (DEP) between DS and HC as well as symptomatic and asymptomatic DS were carried out two-sided. An uncorrected *p*-value of less than 0.05 was considered statistically significant; in the case of multiple comparisons, the Benjamin–Hochberg method for false discovery

rate was applied, with a *p*-value below 0.05 set as a significance threshold.

All obtained DEP were analyzed for enrichment of biological processes, molecular functions, and specific cellular component using the Gene Ontology (GO, accessed 03/24/2024) database³⁵ and investigated for enrichment of biological signaling pathways using the Kyoto Encyclopedia of Genes and Genomes (KEGG, version 109).³⁶ Protein–protein interactions (PPI) between DEP were assessed with the STRING database (version 12.0).³⁴ All proteins measured on the panel were set as background reference.

Feature selection was performed using absolute shrinkage and selection operator (LASSO) regression with an *l*₁ penalty and a regularization parameter lambda set to 0.107 as chosen by cross-validation. For further analysis, a train and test subset were generated using 80% and 20% of the whole data set, respectively. Weights, accounting for the imbalance in diagnosis outcome, were applied. The resulting selection of proteins relevant to predicting the outcome variable was then further used to assess their diagnostic performance via receiver operating characteristics (ROC) analysis.

3 | RESULTS

3.1 | Demographics

In this study, we included 73 adults with DS and 15 euploid HC. There were no significant differences in age or sex distribution (Table 1, both $p > 0.05$). As expected, there was a significant difference between the age of sDS and aDS, with the latter being around 20 years younger ($p < 0.001$). Further, we found a significantly worse performance on the CAMCOG-DS in the symptomatic DS group ($p < 0.001$).

3.2 | DS versus HC

Between DS and HC, we found 253 proteins exhibiting different plasma levels after false discovery rate (FDR) correction. Out of these, 211 were increased (Figure 1A) in DS, while the remaining 42 showed decreased levels in DS compared to HC. Mapping each of these DEP to their originating chromosome, we found 11 proteins to be encoded on chr21, all of them with higher levels in DS (Figure 1B).

To further gain understanding of the relevance of the proteomic differences between both groups for biological and molecular function as well as known signaling pathways in humans, we conducted protein enrichment analysis. Utilizing the KEGG database, we found two significantly enriched processes, namely cytokine–cytokine receptor interaction (hsa04060) and cell adhesion molecules (hsa04514) connected with increased protein levels in DS. When assessing the proteomes with the GO database, we found enriched processes of acute inflammatory response, cell binding and regulatory signaling, and neuronal growth connected to significantly increased protein levels, while significantly reduced protein levels in DS were matched to several pathways relating to the cell cycle, RNA and DNA processing as

TABLE 1 Demographics of all participants included in this study.

Parameter	DS, N = 73	aDS, N = 49	sDS, N = 24	HC, N = 15
Age at visit (years)	38 ± 12	32 ± 8	51 ± 8	44 ± 17
Sex				
Female	34 (47%)	24 (49%)	10 (42%)	9 (60%)
Male	39 (53%)	25 (51%)	14 (58%)	6 (40%)
Leucocyte blood level (μL ⁻¹)		5.7 ± 1.4	5.8 ± 1.4	
NA		6	3	
Diagnosis of AID	49 (67%)	36 (74%)	13 (60%)	
CAMCOG-DS (%)		63 ± 17	46 ± 18	
NA		2	1	
Amyloid (A) status				
Positive		0	12	
Negative		3	0	
NA		46	12	
Tau (T) status				
Positive		0	5	
Negative		1	3	
NA		48	16	
Neurodegeneration (N) status				
Positive		0	10	
Negative		9	1	
NA		40	13	

Abbreviations: aDS, asymptomatic Down syndrome; AID, autoimmune disease; CAMCOG-DS, Cambridge Cognitive Examination for Older Adults with Down Syndrome; DS, Down syndrome; HC, healthy control; NA, not available; sDS, symptomatic Down syndrome.

well as chromatin and DNA integrity, cellular stress, and macromolecular biosynthesis (Figure 1C,D). A comprehensive list of all significant pathway findings between the groups can be found in the supplements (Table S2).

For a better understanding of the functional protein networks affected by the discovered DEP, we assessed PPI with the STRING database³⁴ to visualize and further categorize them into functional clusters. Enrichment within each cluster was further analyzed leveraging the GO, KEGG, and Reactome³⁷ database (Table S3, Figure S1).

DEP between DS and HC were clustered into nine subnetworks with the largest one consisting of 162 DEP showing functional enrichment, among others, for cytokine–cytokine receptor interaction (hsa04060), immune system processes (GO:0002376), transmembrane signaling receptor activity (GO:0004888), and cytokine signaling in the immune system (HSA-1280215). The second largest cluster, made up of 16 DEP, was associated with synaptic function such as PPI at synapses (HSA-6794362) as well as voltage-gated potassium channel complexes (GO:0008076) and gamma-aminobutyric acid (GABA)-ergic synapses (GO:0098982). For another cluster with eight corresponding DEP, we found protein interactions relating to the lipid metabolic pro-

cess (GO:0006629) and phospholipid metabolism (HSA-1483257). Some for the remaining clusters reported associations of protein interactions included azurophile granules (GO:0042582) and lysosomes (GO:0043202, 4 DEP) as well as carbonate dehydratase activity (GO:0004089) and nitrogen metabolism (hsa00910, 2 DEP).

3.3 | sDS versus aDS

In the interest of discovering potential plasma biomarkers for the diagnosis and/or progression of DS-AD, we repeated this analysis in the subgroup of adults with DS, dichotomizing them by their clinical diagnosis of being either symptomatic or asymptomatic. Here, we found 142 DEP after FDR correction. Specifically, 133 of DEP were increased in symptomatic DS, while the remaining nine DEP exhibited decreased levels in this group. Interestingly, none of the DEP originated from chr21 (Figure 2A,B).

When assessing significantly enriched signaling pathways with the KEGG database, we found cytokine–cytokine receptor interaction as significantly enriched in symptomatic DS, in addition to antigen processing and presentation and graft-versus-host-disease. One pathway seemed to be suppressed in the symptomatic phenotype which was the Fc-γ-R-mediated phagocytosis. Within the GO database, symptomatic adults with DS showed significant enrichment in 139 GO terms. Of them, 33 were associated with pathway activation, including immune-related processes such as lymphocyte chemotaxis, major histocompatibility complex (MHC)-protein complex binding, and glial cell proliferation, further signaling processes such as chemokine receptor binding, cytokine activity, glycosaminoglycan binding, and G-protein–coupled receptor binding, as well as growth plasma levels and metabolic activities. Pathways significantly suppressed by means of reduced expression of the corresponding proteins in sDS included cellular structure and organization, cell proliferation, cell signaling and communication, and transcription and translation (Figure 2C,D, Table S4).

Looking at PPI networks within DS, DEP between symptomatic and asymptomatic adults were clustered into 13 functional subnetworks (Figure 3) with the largest consisting of 72 proteins showing enrichment for terms including cytokine–cytokine receptor interaction (hsa04060), signaling (GO:0023052), death receptor activity (GO:0005035), and immune system (HSA-168256). The second largest cluster made up of 10 DEP was associated with extracellular matrix structural constituents (GO:0005201, GO:0031012) and molecules associated with elastic fibers (HSA-2129379). Another cluster counting six DEP exhibited functional association of PPI with enzymatic activity such as protein digestion and absorption (hsa04974) and peptidase (GO:0008233) as well as hydrolase (GO:0016787) activity. Finally, the fourth largest cluster consisted of five DEP associated with the Wnt signaling pathway (hsa04310, GO:0016055). All remaining clusters can be found in the supplements (Table S5).

LASSO feature selection (balanced accuracy: 0.9, sensitivity: 0.8, specificity: 1, positive predictive value: 1, negative predictive value: 0.7143) investigated all proteins measured regarding their potential

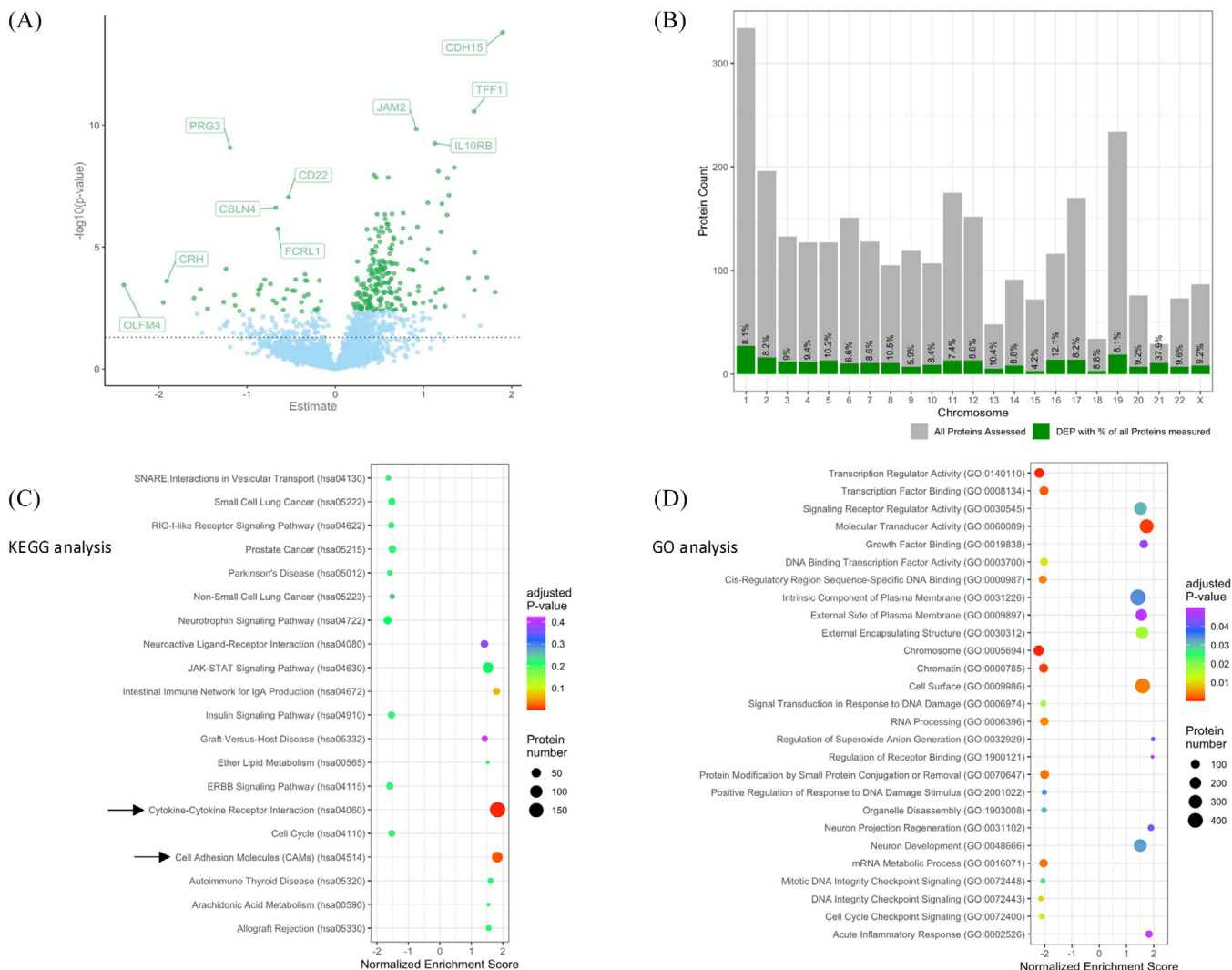


FIGURE 1 (A) Volcano plot displaying the differences of DEP in adults with DS compared to HC. Accordingly, 211 DEP were found to be increased (e.g., CDH15, TFF1, and IL10RB) and 42 DEP showed reduced circulating protein levels in DS versus HC (e.g., OLFM4, PRG3, and CBLN4). (B) Distribution of all DEP (green) in relation to all proteins assessed (grey) and their respective chromosome revealed 11 DEP (37.9%) originating from chr21. (C) Top 20 enriched pathways from KEGG analysis in DS versus HC with the 10 most positive and negative normalized enrichment score (NES), respectively, and where the size of the dot refers to the number of proteins that have been put in for analysis and are annotated in the respective term. Here, cytokine-cytokine receptor interaction and CAMs turned up significantly enriched (arrows). (D) Significantly enriched terms from GO analysis for increased and suppressed pathways in DS compared to HC. DEP, differentially expressed proteins; DS, Down syndrome; GO, Gene Ontology; HC, healthy control; KEGG, Kyoto Encyclopedia of Genes and Genomes.

for predicting the outcome of sDS versus aDS and selected 15 non-zero coefficients, with, apart from NFL and GFAP, 13 further potential markers spanning areas of immune response, neurotransmission, and receptor signaling (Figure 4, Table S6). Eleven proteins contributed positively toward the probability of sDS, while the remaining four exhibited a relevant negative relationship with the outcome of sDS versus aDS (Figure S2).

We assessed the diagnostic performance of all 15 proteins with ROC analysis which revealed an AUC above 0.75 for nine of the LASSO-selected features (Table 2, Figure 5), specifically NFL, GFAP, ectodysplasin A2 receptor (EDA2R), C-X-C motif chemokine 17 (CXCL17) and cluster of differentiation 14 (CD14), insulin-like growth factor bind-

ing protein-2 (IGFBP2), spondin-1 (SPON1), cerebellin 4 (CBLN4), and neuronal-specific septin-3 (SEPTIN3).

4 | DISCUSSION

This cross-sectional study investigated proteomic profiles in adults with and without DS leveraging the Olink Explore 3072.

For 253 DEP between DS and HC, enrichment analysis reported coordinated up-regulation in protein levels for immune activity, cell and cytokine signaling, and neuronal growth while proteins for DNA processing, cell cycle, and biosynthesis were significantly down-regulated

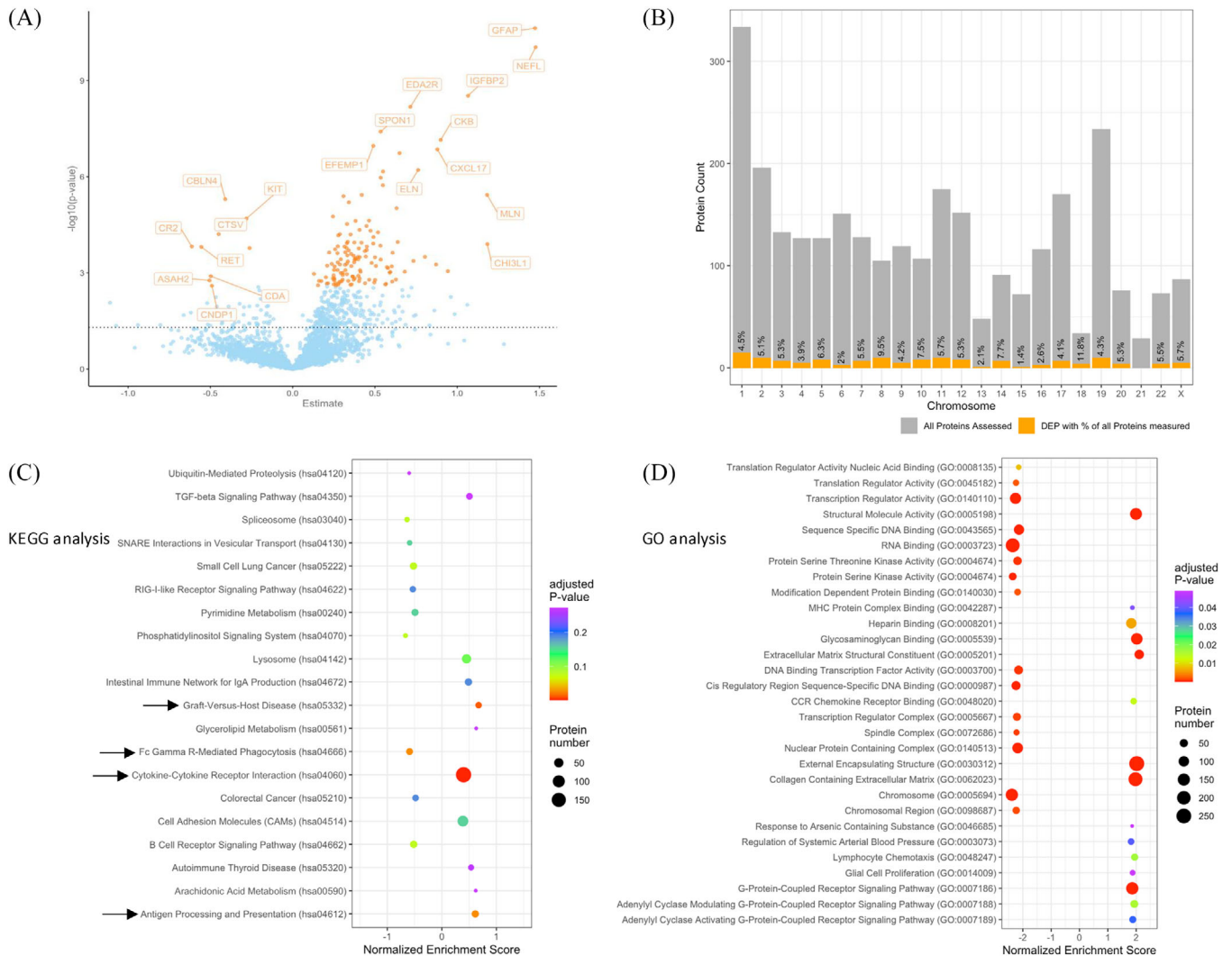


FIGURE 2 (A) Volcano plot displaying the differences of DEP in symptomatic versus asymptomatic adults with DS. Here, 133 DEP were found to with increased levels (e.g., GFAP, NEF, and IGFBP2) and 9 DEP showed decreased protein levels (e.g., CBLN4, CR2, and RET). (B) Distribution of all DEP (orange) in respect to all proteins assessed (grey) between symptomatic and asymptomatic DS regarding their respective chromosome with none of them originating from chr21. (C) Top 20 terms from KEGG Analysis, with 3 of them showing significant positive enrichment for symptomatic DS, and 1 negatively enriched pathway (arrows). (D) Significant terms from GO analysis for top enhanced or diminished pathways, respectively, in symptomatic versus asymptomatic DS. DEP, differentially expressed proteins; DS, Down syndrome; GFAP, glial fibrillary acidic protein; IGFBP2, insulin-like growth factor binding protein-2; KEGG, Kyoto Encyclopedia of Genes and Genomes; NEF, negative regulatory factor.

in DS. Eleven DEP originated from chr21, with reported involvement in immune response regulation, cellular adhesion, and tissue remodeling.^{29,38,39} Analyzing PPI, large clusters related to enhanced processes such as regulation of leucocyte proliferation, cytokine production, synaptic function, and lipid metabolism in DS.

This is in line with observations of a dysregulated and highly active immune system throughout the lifespan of DS,^{40,41} resulting in higher susceptibility for autoimmune diseases,^{42,43} further corroborated by reports of increased cytokine blood levels in children and adults with DS^{40,42,44} and a higher chemokine and cytokine response from monocyte-derived dendritic cells of DS patients.⁴⁴ Also, a study with SomaScan in blood reported similar pathway enrichments of inflammatory response, immune control, and regulation of neurogenesis between DS and HC.²⁹

DS is considered a primary interferonopathy given that increased cytokine signaling is likely due to the fact that four of six interferon (IFN) receptors are encoded on chr21.^{43,45} We, too, found correlates of enhanced signaling in DS with increased levels of IFNAR1, IL10RB, originating chr21, as well as IFNGR1, ILR1, IL4R, and IL20RB. Since chemokines and cytokines play a vital role by regulating cell constitution, migration, and balancing pro- and anti-inflammatory processes,^{29,46} dysregulations could lead to progressive neuroinflammation contributing to DS-AD.^{26,27} Further, dysregulated cell adhesion molecules pathways associated with chr21 in DS versus HC could affect cellular interactions and tissue remodeling. Junctional adhesion molecule 2 (JAM2) is involved in endothelial tight junction formation,³⁸ has been associated with AD,⁴⁷ and could potentially affect the blood-brain barrier permeability in DS-AD. Elevated levels of

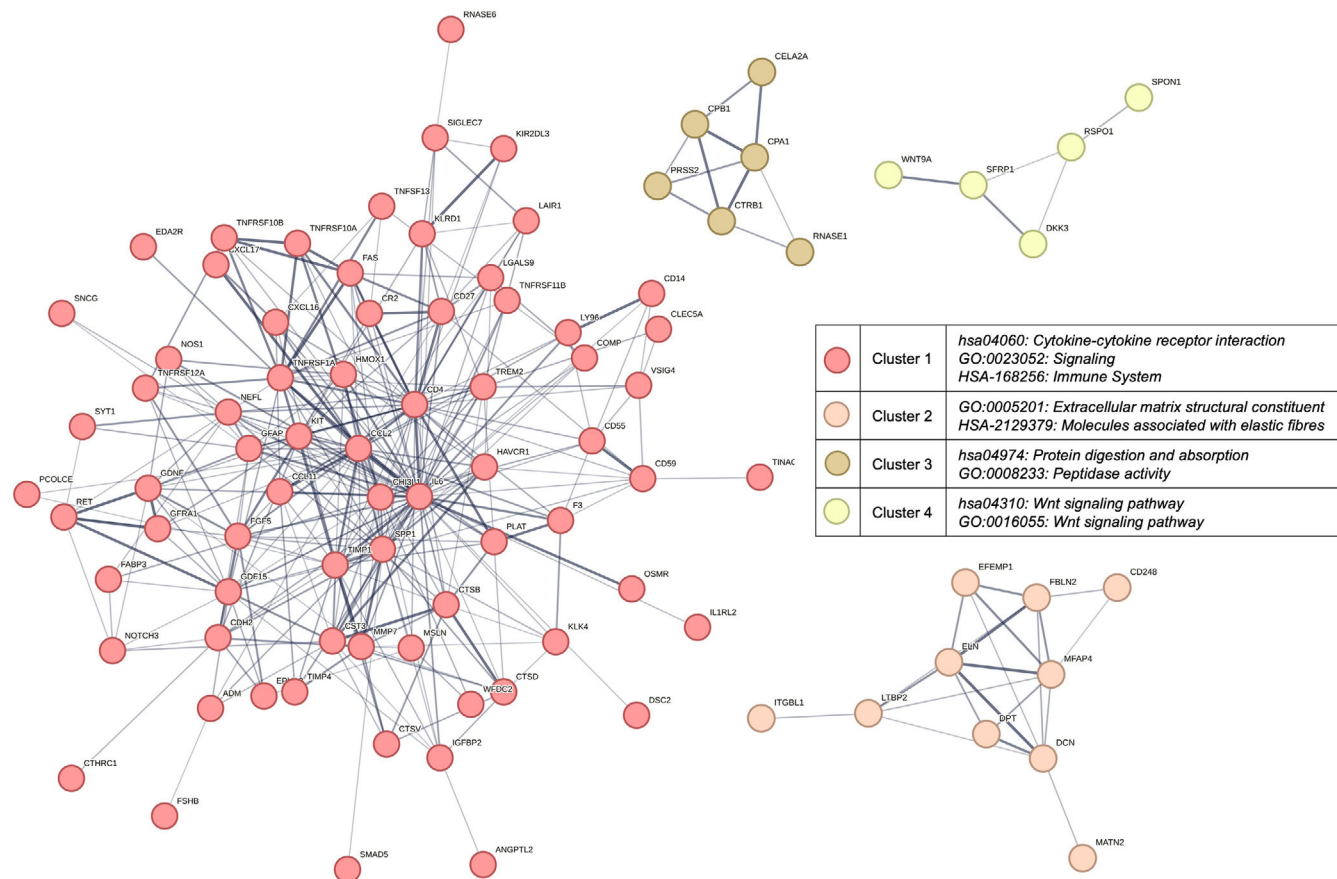


FIGURE 3 Top four clusters using the Markov–Cluster algorithm from PPI STRING network analysis of DEP between symptomatic and asymptomatic adults with DS with most significant enriched terms annotated in the legend suggesting differences in immune system signaling (cluster 1), the makeup of the extracellular matrix (cluster 2), enzymatic protein digestion (cluster 3), and Wnt signaling (cluster 4). Each node represents one DEP while each edge represents functional interaction between two DEP with the thickness of the edge increasing with the confidence of the proposed shared function. DEP, differentially expressed proteins; DS, Down syndrome; PPI, protein–protein interactions.

neural cell adhesion molecule 2 (NCAM2) and Collagen Type XVIII Alpha 1 Chain (COL18A1) arguably contribute to the intellectual disability phenotype³⁹ and decreased risk for solid tumors⁴⁸ in DS, respectively. However, their impact within DS-AD remains unclear. Increased levels of cystatin B (CSTB) though have been found in DS-AD brains, potentially affecting amyloid metabolism and neuroinflammatory processes via modulation of cysteine cathepsins.⁴⁹

Pathways of neuronal development and regeneration were also significantly enriched in DS. Studies in induced pluripotent DS stem cells suggest abnormal neuronal differentiation affecting neuronal architecture and density as well as a number and length of neurites, and differentiation into GFAP-positive cells, thereby promoting a shift from the neuronal to the astroglial and oligodendroglial phenotype,^{50,51} which could facilitate later developments of neuroinflammation and AD pathology.

Pathway analyses in blood from euploid sAD versus HC leveraging OLINK reported changes in signal transduction, apoptosis and inflammatory pathways, cell proliferation, and monocyte chemotaxis,⁵² as well as positive regulation of protein kinase B signaling, growth factor binding, and cytokine–cytokine receptor interaction.⁵³ While the lat-

ter indicates some shared proteomic alterations with our analysis, most of the reported pathways suggest distinct differences in the proteomic dynamics between sAD and DS-AD, underlining the value of DS as a unique model for AD.

Subsequently, we identified 142 DEP between sDS and aDS, none originating from chr21, suggesting proteomic differences not directly resulting from increased gene dosage of chr21, but rather of downstream effects.

The major findings in enrichment analysis within DS concerned dysregulation of immune function and cell signaling, tissue differentiation, extracellular constituents, and metabolic activity, which was mirrored in PPI analysis.

Dysregulated cytokine signaling, antigen presentation, and signaling involving G-protein coupled receptors suggest imbalances in immune cell communication and glial cell proliferation in sDS, in line with an overactive immune environment by further amplifying inflammatory responses, possibly resulting from AD pathology.⁵⁴ Studies report dynamic cytokine profiles along the DS-AD spectrum, resembling an early and a late neuroinflammatory period, with early peaks of upregulated pro-inflammatory markers like interleukin (IL)-6, IL-1 β ,

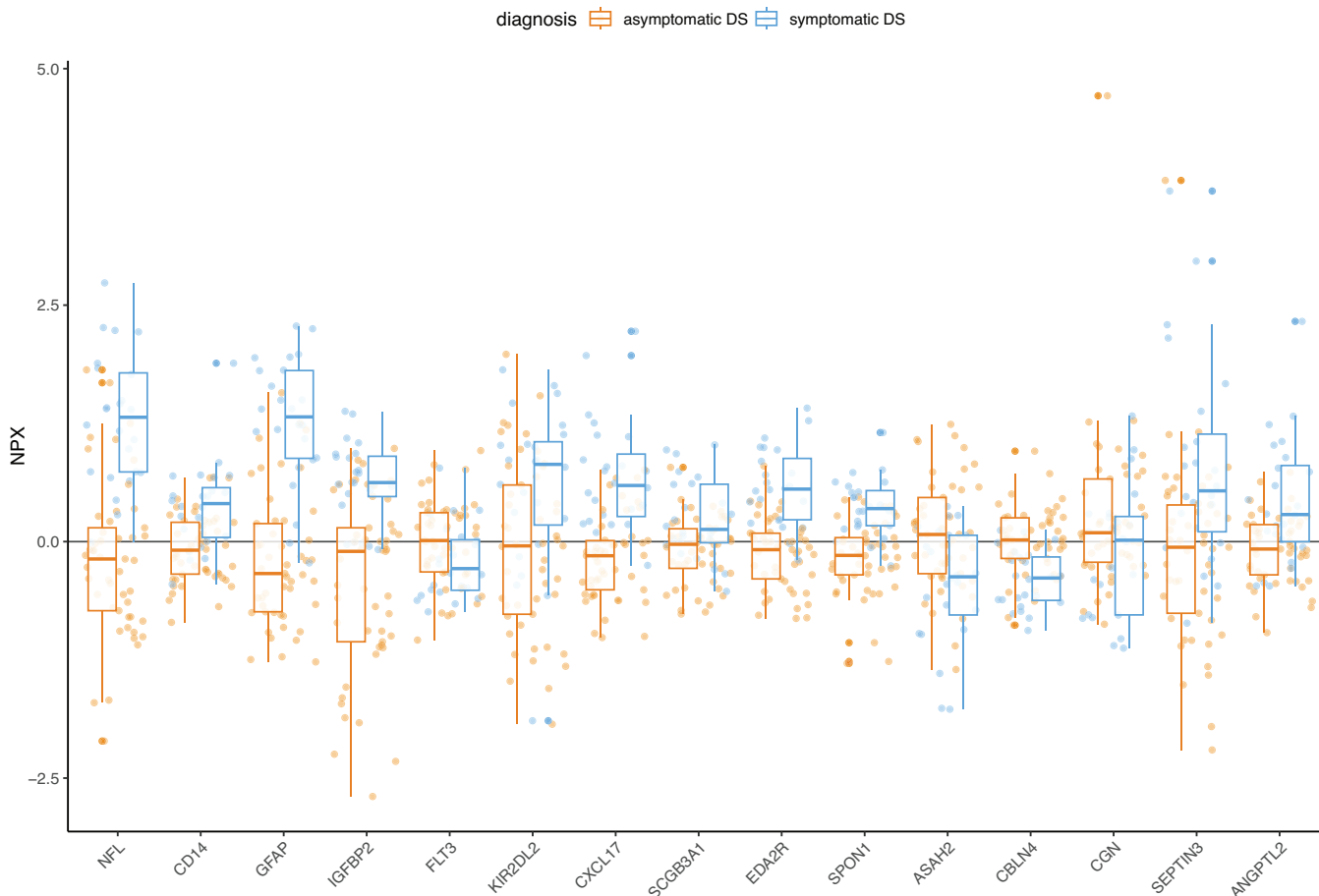


FIGURE 4 Box plots of individual NPX levels from symptomatic and asymptomatic adults with Down syndrome for all 15 proteins received from LASSO feature selection. LASSO, least absolute shrinkage and selection operator; NPX, normalized protein expression.

IL-15, and macrophage-derived chemokine (MDC) around age 20.²⁶ Likewise, elevated plasma levels of sTREM2 have been observed in young DS.⁵⁵ Late inflammatory processes, however, have been characterized as persistent activation of increasingly dystrophic microglia and decreased immune markers like IL-10, IL-12, tumor necrosis factor- α (TNF α), and IFN γ , while levels of IL-6, IL-8, IL-15, and monocyte chemoattractant protein-1 (MCP-1) increased.²⁶ A phenotype of microglial activation characterized by elevation of CD86, IL-10, TNF α , IL-1 β , IL-6, and decrease in IL-12, already in asymptomatic patients below age 40, has been reported in DS brain tissue and shows further exacerbation with age²⁷ while investigations in blood in DS with or without AD showed elevated levels of proinflammatory markers with a combined measure of amyloid- β and inflammatory agents predicting future cognitive decline the following 24 months.⁵⁶

The downregulation of several proteins in sDS relates to considerable cell cycle dysregulation suggestive of altered cellular growth, repair mechanisms, cell stability, and chromosomal segregation. Alterations in cell cycle and proliferation potency have been reported in trisomic cells,^{57,58} and a DS mouse model showed decelerated cell proliferation with increasing age⁵⁹ while ribosomal biogenesis exhibited dysregulation in older compared to younger DS individuals.⁶⁰ Interest-

ingly, a study reported that increases in APP expression could inhibit cell proliferation by regulating global gene expression.⁶¹ Since the onset of symptomatic DS-AD is inevitably intertwined with accumulating AD pathology and therefore increasing age, our findings support potential molecular changes during the life span of DS possibly drive aging processes and benefit AD pathophysiology.

Applying LASSO, we identified 15 proteins for the distinction between sDS and aDS, 9 of which were identified as DEP and exhibited an AUC above 0.75.

NfL blood levels increase early in DS-AD, with baseline concentrations predicting dementia status and preceding amyloid PET changes by up to 10 years^{14,15,62,63} while GFAP, rises significantly in prodromal and symptomatic DS-AD, correlating well with imaging measures of AD (-related) pathology.²⁴ We identified both as relevant features for sDS, confirming strong diagnostic performance (AUC > 0.9).^{15,24}

EDA2R is enriched in reactive astrocytes^{64,65} and higher levels have been associated with smaller grey matter volume and impaired fluid cognitive ability in euploids.⁶⁶ In our sDS cohort, EDA2R overexpression could therefore contribute to cognitive decline via chronic reactive astroglia.

The astrocytic signaling protein IGFBP2 is increased in AD,⁶⁷ associated with clinical diagnosis of mild cognitive impairment or dementia,

Top LASSO selected Features (AUC>0.75) predicting sDS

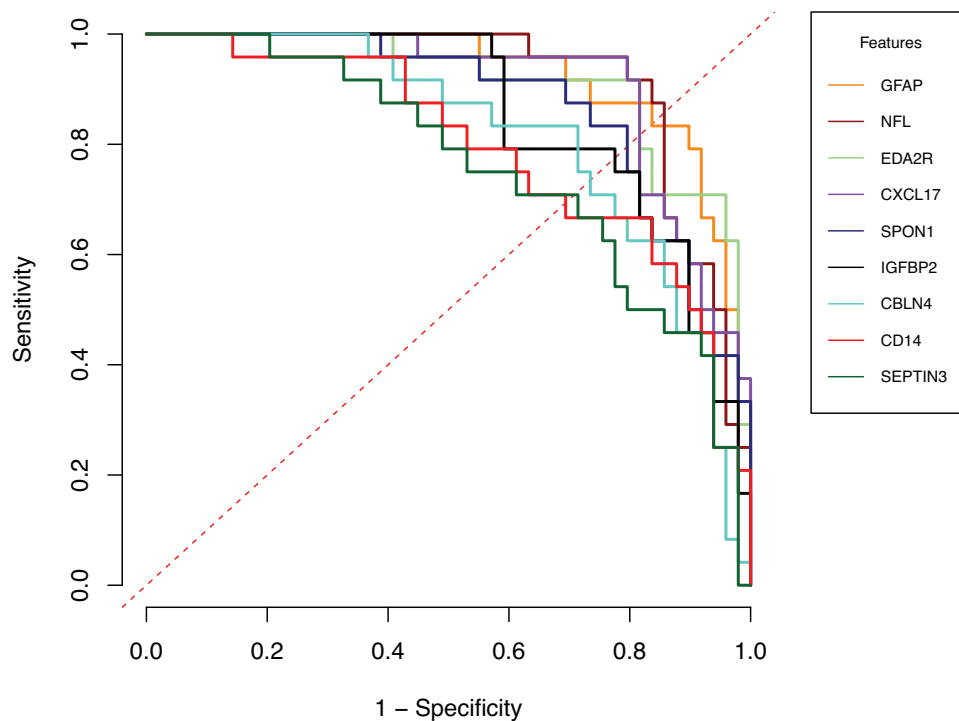


FIGURE 5 All nine proteins received from feature selection analysis with their corresponding AUC for diagnostic performance of differentiating between symptomatic and asymptomatic DS with corresponding ROC curves and an AUC over 0.75. AUC, area under the curve; DS, Down syndrome; ROC, receiver operating characteristics.

AD-like brain atrophy, and CSF tau levels,^{68,69} supporting our finding of significantly increased blood levels in sDS.

Soluble CD14 has been shown to enhance immune response to lipopolysaccharides in human euploid cells⁷⁰ and is increased in sDS, suggesting a further exacerbation of the augmented reaction to lipopolysaccharides reported in DS by CD14-positive-monocytes-derived cells.⁴⁴ We, too, found it significantly increased in sDS, potentially contributing to dysregulated immune-related pathways, exhibiting a moderate association with disease status.

While CXCL17 is mostly associated with chemotaxis in mucosal tissue,⁷¹ a mouse model study found increased cerebellar expression of CXCL17 upon stimulation by peripheral inflammatory agents,⁷² which could explain the elevated levels and good discriminative power within our analysis.

SPON1, significantly increased in sDS, is reported to prohibit enzymatic release of amyloid by binding to APP,⁷³ thereby possibly mediating cognitive decline and structural brain changes in AD,^{74,75} and increase in CSF in autosomal-dominant AD, as early as 30 years prior symptom onset.⁷⁶

Finally, synaptic proteins SEPTIN3 and CBLN4 were increased and decreased, respectively, in sDS, hinting at compensatory processes ameliorating AD-related synaptic dysfunction. Indeed, disorganization and accumulation of SEPTIN3 has been associated with complement-dependent synapse loss in AD⁷⁷ and its gene polymorphisms are considered relevant for AD pathology susceptibility.⁷⁸ Conversely,

its elevated levels in sDS could also hint at increased release from perishing synapses rather than overexpression, as proposed with beta-synuclein.²⁵ Meanwhile, CBLN4, pivotal for synaptic formation and maintenance, has been suggested as a treatment target of AD due to its amelioration of amyloid-related synaptic dysfunction.⁷⁹ Therefore, reduced expression levels in sDS might be contributing to synaptic dysfunction.

Our study has certain limitations. Our sample size of 73 adults with DS and 15 HC is rather small, warranting further analyses with larger cohorts to validate present findings. Secondly, our cross-sectional sample allows only limited interpretation of the potential biomarkers uncovered; however, DS-AD arguably allows for a pseudo-longitudinal interpretation similar to autosomal-dominant AD⁸⁰ due to the genetically determined course of pathology.⁸¹ Further, lack of comprehensive data for amyloid, tau, and markers of neurodegeneration for the majority of the cohort prohibited us from further differentiating potential biomarkers according to neuropathological status. Finally, LASSO analysis was not adjusted for age due to its high correlation with diagnosis (Pearson $r = 0.74$) and strong potential for solely predicting diagnosis, indicating a highly significant association (logistic regression estimate = 0.225, $p < 2e-16$, Akaike information criterion [AIC] = 124,231). While this approach avoids conflating age-related effects with the outcome of diagnosis, it limits our ability to fully disentangle their independent contributions, which we plan to address in future analyses.

TABLE 2 All 15 proteins received from feature selection analysis with their corresponding AUC for diagnostic performance of differentiating between symptomatic and asymptomatic DS.

Feature	AUC
GFAP	0.925
NFL	0.916
EDA2R	0.91
CXCL17	0.904
SPON1	0.872
IGFBP2	0.855
CBLN4	0.815
CD14	0.792
SEPTIN3	0.759
ANGPTL2	0.732
KIR2DL2	0.72
ASAH2	0.711
SCGB3A1	0.71
FLT3	0.654
CGN	0.617

Abbreviation: AUC, area under the curve; DS, Down syndrome.

In conclusion, this work confirmed NFL and GFAP as powerful blood markers for DS-AD and further identified several novel potential biomarkers with considerable power for diagnosing DS-AD. Motivated by the need for the implementation of an easily accessible and well-tolerated alternative to lumbar punctures and imaging protocols, the identified candidates warrant further investigation.

ACKNOWLEDGMENTS

The authors acknowledge the technical support of the Metabolomics and Proteomics Core Facility at Helmholtz Munich as well as the support from the LMU Department of Statistics (StaBLab). This work was funded by the Alzheimer Forschung Initiative (#22049). Further support was received by the Deutsche Forschungsgemeinschaft (DFG, German Research Foundation) under Germany's Excellence Strategy within the framework of the Munich Cluster for Systems Neurology (EXC 2145 SyNergy - ID 390857198), by the Bundesministerium für Bildung und Forschung (BMBF) project CLINSPECT-M (FKZ: FKZ161L0214B, FKZ161L0214C, FKZ16LW0248), by the Else Kröner-Fresenius-Stiftung (2020_EKEA.09), by the Jerome LeJeune Foundation and the Verum Foundation. F.J.M.M. received support from Project PID2022-137629OA-I00 and grant RYC2021-030875-I, funded by MICIU/AEI/10.13039/501100011033 and respectively by ERDF/EU and European Union NextGenerationEU/PRTR. This work was also supported by the Fondo de Investigaciones Sanitarias, Carlos III Health Institute (INT21/00073, PI20/01473 and PI23/01786 to J.F.) and the Centro de Investigación Biomédica en Red sobre Enfermedades Neurodegenerativas Program 1, partly jointly funded by Fondo Europeo de Desarrollo Regional, Unión Europea, Una Man-

era de Hacer Europa, by the National Institutes of Health grants (R01 AG056850; R21 AG056974, R01 AG061566, R01 AG081394, and R61AG066543 to J.F.), the Department de Salut de la Generalitat de Catalunya, Pla Estratègic de Recerca i Innovació en Salut (SLT006/17/00119 to J.F.) and the Fundació Tatiana Pérez de Guzmán el Bueno (IIBSP-DOW-2020-151 o J.F.) as well as the Horizon 2020-Research and Innovation Framework Programme from the European Union (H2020-SC1-BHC-2018-2020 to J.F.).

CONFLICT OF INTEREST STATEMENT

J.F. reports receiving personal fees for service on the advisory boards, adjudication committees or speaker honoraria from AC Immune, Adamed, Alzheon, Biogen, Eisai, Esteve, Fujirebio, Ionis, Laboratorios Carnot, Life Molecular Imaging, Lilly, Lundbeck, Perha, Roche and outside the submitted work as well as holding a patent for markers of synaptopathy in neurodegenerative disease (licensed to Adx, EPI8382175.0). G.H. serves as a consultant for Abbvie, Alzprotect, Amylyx, Aprinoia, Asceneuron, Bayer, Bial, Biogen, Biohaven, Epidarex, Ferrer, Kyowa Kirin, Lundbeck, Novartis, Retrotope, Roche, Sanofi, Servier, Takeda, Teva, UCB; received honoraria for scientific presentations from Abbvie, Bayer, Bial, Biogen, Bristol Myers Squibb, Esteve, Kyowa Kirin, Pfizer, Roche, Teva, UCB, Zambon; holds patents (US 10,918,628 B2; EP 17 787 904.6-1109/3 525 788); received publication royalties from Academic Press, Kohlhammer, and Thieme. J.L. reports speaker fees from Bayer Vital, Biogen, EISAI, TEVA, Zambon, Esteve, Merck and Roche, consulting fees from Axon Neuroscience, EISAI, and Biogen, author fees from Thieme medical publishers and W. Kohlhammer GmbH medical publishers and is inventor in a patent "Oral Phenylbutyrate for Treatment of Human 4-Repeat Tauopathies" (PCT/EP2024/053388) filed by LMU Munich. In addition, he reports compensation for serving as chief medical officer for MODAG GmbH, is beneficiary of the phantom share program of MODAG GmbH and is inventor in a patent "Pharmaceutical Composition and Methods of Use" (EP 22 159 408.8) filed by MODAG GmbH, all activities outside the submitted work. J.M.G., S.M.H., S.K., C.J.M., G.N., S.V.L., F.J.M.M., A.P., C.P., R.R.Z., A.S., K.S., E.W., and O.W. report no conflict of interest. Author disclosures are available in the [supporting information](#).

CONSENT STATEMENT

All DS patients included in this analysis were recruited through our AD21 cohort study investigating AD in adults with DS; each individual or their respective legal proxy provided informed written consent prior to inclusion. Blood samples of healthy controls without the diagnosis of DS from our biobank were also obtained after providing written consent. Both studies are supervised and approved by the LMU ethics committee (DS: #17-126, HC: #20-0997) and conducted in accordance with the Declaration of Helsinki.

ORCID

Olivia Wagemann  <https://orcid.org/0000-0003-3211-9105>

Johannes Levin  <https://orcid.org/0000-0001-5092-4306>

REFERENCES

- Jack CR, Bennett DA, Blennow K, et al. NIA-AA research framework: toward a biological definition of Alzheimer's disease. *Alzheimers Dement*. 2018;14(4):535-562. doi:10.1016/j.jalz.2018.02.018
- Dubois B, Villain N, Frisoni GB, et al. Clinical diagnosis of Alzheimer's disease: recommendations of the International Working Group. *Lancet Neurol*. 2021;20(6):484-496. doi:10.1016/S1474-4422(21)00066-1
- Liu P, Wang Y, Sun Y, Peng G. Neuroinflammation as a potential therapeutic target in Alzheimer's disease. *Clin Interv Aging*. 2022;17:665-674. doi:10.2147/CIA.S357558
- Tönnies E, Trushina E. Oxidative stress, synaptic dysfunction, and Alzheimer's disease. *J Alzheimers Dis*. 2017;57(4):1105-1121. doi:10.3233/JAD-161088
- Fortea J, Zaman SH, Hartley S, Rafii MS, Head E, Carmona-Iragui M. Alzheimer's disease associated with Down syndrome: a genetic form of dementia. *Lancet Neurol*. 2021;20:930-942. doi:10.1016/S1474-4422(21)00245-3
- Gyure KA, Durham R, Stewart WF, Smialek JE, Troncoso JC. Intraneuronal A β -amyloid precedes development of amyloid plaques in Down syndrome. *Arch Pathol Lab Med*. 2001;125(4):489-492. doi:10.5858/2001-125-0489-IAAPDO
- Wiseman FK, Alford KA, Tybulewicz VL, Fisher EM. Down syndrome—recent progress and future prospects. *Hum Mol Genet*. 2009;18(R1):R75-R83. doi:10.1093/hmg/ddp010
- Loosli SV, Neumann LC, Wlasich E, et al. Measurement properties of the German version of the Cambridge examination for mental disorders of older people with Down syndrome and others with intellectual disabilities (CAMDEX-DS). *J Intellect Dev Disabil*. 2024;49(4):402-414. doi:10.3109/13668250.2024.2317794
- Nübling G, Wagemann O, Loosli SV, Wlasich E, Danek A, Levin J. [Diagnosing Alzheimer dementia in people with Down syndrome in accordance with ICD, DSM V and A/T/N system]. *Fortschr Neurol Psychiatr*. 2022;90(9):416-420. doi:10.1055/a-1771-6273
- Deb S, Hare M, Prior L. Symptoms of dementia among adults with Down's syndrome: a qualitative study. *J Intellect Disabil Res*. 2007;51(Pt 9):726-739. doi:10.1111/j.1365-2788.2007.00956.x
- Smith E, Hedge C, Jarrold C. A novel framework to measure executive function in down syndrome with applications for early clinical diagnosis of dementia. *Am J Intellect Dev Disabil*. 2019;124(4):354-373. doi:10.1352/1944-7558-124.4.354
- Nieuwenhuis-Mark RE. Diagnosing Alzheimer's dementia in Down syndrome: problems and possible solutions. *Res Dev Disabil*. 2009;30(5):827-838. doi:10.1016/j.ridd.2009.01.010
- Petersen ME, O'Bryant SE. Blood-based biomarkers for Down syndrome and Alzheimer's disease: a systematic review. *Dev Neurobiol*. 2019;79(7):699-710. doi:10.1002/dneu.22714
- Fortea J, Vilaplana E, Carmona-Iragui M, et al. Clinical and biomarker changes of Alzheimer's disease in adults with Down syndrome: a cross-sectional study. *Lancet*. 2020;395(10242):1988-1997. doi:10.1016/S0140-6736(20)30689-9
- Fortea J, Carmona-Iragui M, Benejam B, et al. Plasma and CSF biomarkers for the diagnosis of Alzheimer's disease in adults with Down syndrome: a cross-sectional study. *Lancet Neurol*. 2018;17(10):860-869. doi:10.1016/S1474-4422(18)30285-0
- Schupf N, Patel B, Pang D, et al. Elevated plasma β -amyloid peptide A β 42 levels, incident dementia, and mortality in Down syndrome. *Arch Neurol*. 2007;64(7):1007. doi:10.1001/archneur.64.7.1007
- Coppus AMW, Schuur M, Vergeer J, et al. Plasma β amyloid and the risk of Alzheimer's disease in Down syndrome. *Neurobiol Aging*. 2012;33(9):1988-1994. doi:10.1016/j.neurobiolaging.2011.08.007
- Alhajraf F, Ness D, Hye A, Strydom A. Plasma amyloid and tau as dementia biomarkers in Down syndrome: systematic review and meta-analyses. *Dev Neurobiol*. 2019;79(7):684-698. doi:10.1002/dneu.22715
- Lleó A, Zetterberg H, Pegueroles J, et al. Phosphorylated tau181 in plasma as a potential biomarker for Alzheimer's disease in adults with Down syndrome. *Nat Commun*. 2021;12(1):4304. doi:10.1038/s41467-021-24319-x
- Janelidze S, Berron D, Smith R, et al. Associations of plasma Phospho-Tau217 levels with tau positron emission tomography in early Alzheimer disease. *JAMA Neurol*. 2021;78(2):149. doi:10.1001/jamaneurol.2020.4201
- Janelidze S, Christian BT, Price J, et al. Detection of brain Tau pathology in Down syndrome using plasma biomarkers. *JAMA Neurol*. 2022;79(8):797-807. doi:10.1001/jamaneurol.2022.1740
- Schworer EK, Handen BL, Petersen M, et al. Cognitive and functional performance and plasma biomarkers of early Alzheimer's disease in Down syndrome. *Alzheimers Dement (Amst)*. 2024;16(2):e12582. doi:10.1002/dad2.12582
- Petersen ME, Rafii MS, Zhang F, et al. Plasma total-Tau and neurofilament light chain as diagnostic biomarkers of Alzheimer's disease dementia and mild cognitive impairment in adults with Down syndrome. *J Alzheimers Dis*. 2021;79(2):671-681. doi:10.3233/JAD-201167
- Montoliu-Gaya L, Alcolea D, Ashton NJ, et al. Plasma and cerebrospinal fluid glial fibrillary acidic protein levels in adults with Down syndrome: a longitudinal cohort study. *eBioMedicine*. 2023;90:104547. doi:10.1016/j.ebiom.2023.104547
- Oeckl P, Wagemann O, Halbgebauer S, et al. Serum BETA-SYNUCLEIN is higher in Down syndrome and precedes rise of PTAU181. *Ann Neurol*. 2022;92(1):6-10. doi:10.1002/ana.26360
- Flores-Aguilar L, Lulita MF, Kovacs O, et al. Evolution of neuroinflammation across the lifespan of individuals with Down syndrome. *Brain*. 2020;143(12):3653-3671. doi:10.1093/brain/awaa326
- Wilcock DM, Hurban J, Helman AM, et al. Down syndrome individuals with Alzheimer's disease have a distinct neuroinflammatory phenotype compared to sporadic Alzheimer's disease. *Neurobiol Aging*. 2015;36(9):2468-2474. doi:10.1016/j.neurobiolaging.2015.05.016
- Wiseman FK, Pulford LJ, Barkus C, et al. Trisomy of human chromosome 21 enhances amyloid- β deposition independently of an extra copy of APP. *Brain*. 2018;141(8):2457-2474. doi:10.1093/brain/awy159
- Sullivan KD, Evans D, Pandey A, et al. Trisomy 21 causes changes in the circulating proteome indicative of chronic autoinflammation. *Sci Rep*. 2017;7(1):14818. doi:10.1038/s41598-017-13858-3
- Petersen ME, Zhang F, Schupf N, et al. Proteomic profiles for Alzheimer's disease and mild cognitive impairment among adults with Down syndrome spanning serum and plasma: an Alzheimer's Biomarker Consortium-Down Syndrome (ABC-DS) study. *Alzheimers Dement (Amst)*. 2020;12(1):e12039. doi:10.1002/dad2.12039
- American Psychological Association. Neurodevelopmental Disorders; 2022. https://doi.org/10.1176/appi.books.9780890425787.x01_Neurodevelopmental_Disorders </bib >
- Nübling G, Loosli SV, Wlasich E, et al. [A German version of the Cambridge examination for mental disorders of older people with Down's syndrome and others with intellectual disabilities : a diagnostic procedure for detecting dementia in people with Down's syndrome]. *Z Gerontol Geriatr*. 2020;53(6):546-551. doi:10.1007/s00391-019-01591-7
- <https://olink.com/products-services/explore/>
- <https://string-db.org>
- Gene Ontology Consortium. The Gene Ontology (GO) database and informatics resource. *Nucleic Acids Res*. 2004;32(90001):D258-D261. doi:10.1093/nar/gkh036
- Kanehisa M, Furumichi M, Tanabe M, Sato Y, Morishima K. KEGG: new perspectives on genomes, pathways, diseases and drugs. *Nucleic Acids Res*. 2017;45(D1):D353-D361. doi:10.1093/nar/gkw1092

37. Milacic M, Beavers D, Conley P, et al. The reactome pathway knowledgebase 2024. *Nucleic Acids Res.* 2024;52(D1):D672-D678. doi:10.1093/nar/gkad1025
38. Kadry H, Noorani B, Cucullo L. A blood-brain barrier overview on structure, function, impairment, and biomarkers of integrity. *Fluids Barriers CNS.* 2020;17(1):69. doi:10.1186/s12987-020-00230-3
39. Parcerisas A, Ortega-Gascó A, Pujadas L, Soriano E. The hidden side of NCAM family: nCAM2, a key cytoskeleton organization molecule regulating multiple neural functions. *Int J Mol Sci.* 2021;22(18):10021. doi:10.3390/ijms221810021
40. Zhang Y, Che M, Yuan J, et al. Aberrations in circulating inflammatory cytokine levels in patients with Down syndrome: a meta-analysis. *Oncotarget.* 2017;8(48):84489-84496. doi:10.18632/oncotarget.21060
41. Huggard D, Kelly L, Ryan E, et al. Increased systemic inflammation in children with Down syndrome. *Cytokine.* 2020;127:154938. doi:10.1016/j.cyto.2019.154938
42. Malle L, Patel RS, Martin-Fernandez M, et al. Autoimmunity in Down's syndrome via cytokines, CD4 T cells and CD11c+ B cells. *Nature.* 2023;615(7951):305-314. doi:10.1038/s41586-023-05736-y
43. Araya P, Waugh KA, Sullivan KD, et al. Trisomy 21 dysregulates T cell lineages toward an autoimmunity-prone state associated with interferon hyperactivity. *Proc Natl Acad Sci U S A.* 2019;116(48):24231-24241. doi:10.1073/pnas.1908129116
44. Nakashima K, Imai T, Shiraishi A, et al. The immunoreactive signature of monocyte-derived dendritic cells from patients with Down syndrome. *Clin Exp Immunol.* 2024;217(3):291-299. doi:10.1093/cei/uxae048
45. Sullivan KD, Lewis HC, Hill AA, et al. Trisomy 21 consistently activates the interferon response. *eLife.* 2016;5:e16220. doi:10.7554/eLife.16220
46. Hughes CE, Nibbs RJB. A guide to chemokines and their receptors. *FEBS J.* 2018;285(16):2944-2971. doi:10.1111/febs.14466
47. Khondoker M, Newhouse S, Westman E, et al. Linking genetics of brain changes to Alzheimer's disease: sparse whole genome association scan of regional MRI volumes in the ADNI and AddNeuroMed cohorts. *J Alzheimers Dis.* 2015;45(3):851-864. doi:10.3233/JAD-142214
48. Osuna-Marco MP, López-Barahona M, López-Ibor B, Tejera ÁM. Ten reasons why people with down syndrome are protected from the development of most solid tumors - a review. *Front Genet.* 2021;12:749480. doi:10.3389/fgene.2021.749480
49. Wu Y, Mumford P, Noy S, et al. Cathepsin B abundance, activity and microglial localisation in Alzheimer's disease-Down syndrome and early onset Alzheimer's disease; the role of elevated cystatin B. *Acta Neuropathol Commun.* 2023;11(1):132. doi:10.1186/s40478-023-01632-8
50. Guidi S, Bonasoni P, Ceccarelli C, et al. Research Article: neurogenesis impairment and increased cell death reduce total neuron number in the hippocampal region of fetuses with down syndrome. *Brain Pathol.* 2008;18(2):180-197. doi:10.1111/j.1750-3639.2007.00113.x
51. Hibaoui Y, Grad I, Letourneau A, et al. Modelling and rescuing neurodevelopmental defect of Down syndrome using induced pluripotent stem cells from monozygotic twins discordant for trisomy 21. *EMBO Mol Med.* 2014;6(2):259-277. doi:10.1002/emmm.201302848
52. Jiang Y, Zhou X, Ip FC, et al. Large-scale plasma proteomic profiling identifies a high-performance biomarker panel for Alzheimer's disease screening and staging. *Alzheimers Dement.* 2022;18(1):88-102. doi:10.1002/alz.12369
53. Yang S, Ye Z, He P, et al. Plasma proteomics for risk prediction of Alzheimer's disease in the general population. *Aging Cell.* 2024;23(12):e14330. doi:10.1111/acer.14330
54. Gensous N, Bacalini MG, Franceschi C, Garagnani P. Down syndrome, accelerated aging and immunosenescence. *Semin Immunopathol.* 2020;42(5):635-645. doi:10.1007/s00281-020-00804-1
55. Weber GE, Koenig KA, Khrestian M, et al. An altered relationship between soluble TREM2 and inflammatory markers in young adults with Down syndrome: a preliminary report. *J Immunol.* 2020;204(5):1111-1118. doi:10.4049/jimmunol.1901166
56. Lulita MF, Ower A, Barone C, et al. An inflammatory and trophic disconnect biomarker profile revealed in Down syndrome plasma: relation to cognitive decline and longitudinal evaluation. *Alzheimers Dement.* 2016;12(11):1132-1148. doi:10.1016/j.jalz.2016.05.001
57. Antonarakis SE, Skotko BG, Rafii MS, et al. Down syndrome. *Nat Rev Primer.* 2020;6(1):9. doi:10.1038/s41572-019-0143-7
58. Contestabile A, Fila T, Ceccarelli C, et al. Cell cycle alteration and decreased cell proliferation in the hippocampal dentate gyrus and in the neocortical germinal matrix of fetuses with down syndrome and in Ts65Dn mice. *Hippocampus.* 2007;17(8):665-678. doi:10.1002/hipo.20308
59. Rueda N, Flórez J, Martínez-Cué C. Mouse models of Down syndrome as a tool to unravel the causes of mental disabilities. *Neural Plast.* 2012;2012:584071. doi:10.1155/2012/584071
60. Ravaioi F, Zampieri M, Morandi L, et al. DNA methylation analysis of ribosomal DNA in adults with Down syndrome. *Front Genet.* 2022;13:792165. doi:10.3389/fgene.2022.792165
61. Wu Y, Zhang S, Xu Q, et al. Regulation of global gene expression and cell proliferation by APP. *Sci Rep.* 2016;6(1):22460. doi:10.1038/srep22460
62. Strydom A, Heslegrave A, Startin CM, et al. Neurofilament light as a blood biomarker for neurodegeneration in Down syndrome. *Alzheimers Res Ther.* 2018;10(1):39. doi:10.1186/s13195-018-0367-x
63. Ashton NJ, Janelidze S, Al Khleifat A, et al. A multicentre validation study of the diagnostic value of plasma neurofilament light. *Nat Commun.* 2021;12(1):3400. doi:10.1038/s41467-021-23620-z
64. Zamanian JL, Xu L, Foo LC, et al. Genomic analysis of reactive astrogliosis. *J Neurosci.* 2012;32(18):6391-6410. doi:10.1523/JNEUROSCI.6221-11.2012
65. Merienne N, Meunier C, Schneider A, et al. Cell-type-specific gene expression profiling in adult mouse brain reveals normal and disease-state signatures. *Cell Rep.* 2019;26(9):2477.e9-2493.e9. doi:10.1016/j.celrep.2019.02.003
66. Harris SE, Cox SR, Bell S, et al. Neurology-related protein biomarkers are associated with cognitive ability and brain volume in older age. *Nat Commun.* 2020;11(1):800. doi:10.1038/s41467-019-14161-7
67. Lista S, Faltraco F, Prvulovic D, Hampel H. Blood and plasma-based proteomic biomarker research in Alzheimer's disease. *Prog Neurobiol.* 2013;101-102:1-17. doi:10.1016/j.pneurobio.2012.06.007
68. Toledo JB, Da X, Bhatt P, et al. Relationship between plasma analytes and SPARE-AD defined brain Atrophy Patterns in ADNI. Villoslada P, ed. *PLoS ONE.* 2013;8(2):e55531. doi:10.1371/journal.pone.0055531
69. Hu WT, Holtzman DM, Fagan AM, et al. Plasma multianalyte profiling in mild cognitive impairment and Alzheimer disease. *Neurology.* 2012;79(9):897-905. doi:10.1212/WNL.0b013e318266fa70
70. Pahwa R, Devaraj S, Jialal I. The effect of the accessory proteins, soluble CD14 and lipopolysaccharide-binding protein on Toll-like receptor 4 activity in human monocytes and adipocytes. *Int J Obes.* 2016;40(6):907-911. doi:10.1038/ijo.2016.32
71. Gowhari Shabgah A, Jadidi-Niaragh F, Ebrahimzadeh F, et al. A comprehensive review of chemokine CXC17 (VCC1) in cancer, infection, and inflammation. *Cell Biol Int.* 2022;46(10):1557-1570. doi:10.1002/cbin.11846
72. Fil D, Borysiewicz E, Konat GW. A broad upregulation of cerebral chemokine genes by peripherally-generated inflammatory mediators. *Metab Brain Dis.* 2011;26(1):49-59. doi:10.1007/s11011-010-9231-9
73. Ho A, Südhof TC. Binding of F-spondin to amyloid- β precursor protein: a candidate amyloid- β precursor protein ligand that modulates amyloid- β precursor protein cleavage. *Proc Natl Acad Sci U S A.* 2004;101(8):2548-2553. doi:10.1073/pnas.0308655100

74. Park SY, Kang JY, Lee T, Nam D, Jeon CJ, Kim JB. SPON1 can reduce amyloid beta and reverse cognitive impairment and memory dysfunction in Alzheimer's disease mouse model. *Cells*. 2020;9(5):1275. doi:10.3390/cells9051275
75. Jahanshad N, Rajagopalan P, Hua X, et al. Genome-wide scan of healthy human connectome discovers SPON1 gene variant influencing dementia severity. *Proc Natl Acad Sci U S A*. 2013;110(12):4768-4773. doi:10.1073/pnas.1216206110
76. Johnson ECB, Bian S, Haque RU, et al. Cerebrospinal fluid proteomics define the natural history of autosomal dominant Alzheimer's disease. *Nat Med*. 2023;29(8):1979-1988. doi:10.1038/s41591-023-02476-4
77. Györfy BA, Tóth V, Török G, et al. Synaptic mitochondrial dysfunction and septin accumulation are linked to complement-mediated synapse loss in an Alzheimer's disease animal model. *Cell Mol Life Sci*. 2020;77(24):5243-5258. doi:10.1007/s00018-020-03468-0
78. Takehashi M, Alioto T, Stedeford T, et al. Septin 3 gene polymorphism in Alzheimer's disease. *Gene Expr*. 2003;11(5):263-270. doi:10.3727/000000003783992243
79. Chacón PJ, Del Marco Á, Arévalo Á, Domínguez-Giménez P, García-Segura LM, Rodríguez-Tébar A. Cerebellin 4, a synaptic protein, enhances inhibitory activity and resistance of neurons to amyloid- β toxicity. *Neurobiol Aging*. 2015;36(2):1057-1071. doi:10.1016/j.neurobiolaging.2014.11.006
80. Bateman RJ, Xiong C, Benzinger TLS, et al. Clinical and biomarker changes in dominantly inherited Alzheimer's disease. *N Engl J Med*. 2012;367(9):795-804. doi:10.1056/NEJMoa1202753
81. Silverman W, Krinsky-McHale SJ, Kovacs C, et al. Individualized estimated years from onset of Alzheimer's disease - related decline for adults with Down syndrome. *Alzheimers Dement (Amst)*. 2023;15(2):e12444. doi:10.1002/dad2.12444

SUPPORTING INFORMATION

Additional supporting information can be found online in the Supporting Information section at the end of this article.

How to cite this article: Wagemann O, Nübling G, Martínez-Murcia FJ, et al. Exploratory analysis of the proteomic profile in plasma in adults with Down syndrome in the context of Alzheimer's disease. *Alzheimer's Dement*. 2025;21:e70040. <https://doi.org/10.1002/alz.70040>